

Determinazione del PFOS in biota tramite metodica QUECHERs e HPLC/HRMS:

Descrizione e validazione del metodo

Michele Mazzetti

ARPAT - Agenzia Regionale per la Protezione Ambientale della Toscana

OUTLINE

- Introduction
- PFAS Methods – solid matrices
- QuEChERS for PFAS
- Method Performance
- Conclusion

Fluorinated organic compound: Background history

Industrial application of fluorinated organic compounds started (Chlorofluorocarbons (CFC) as refrigerants).



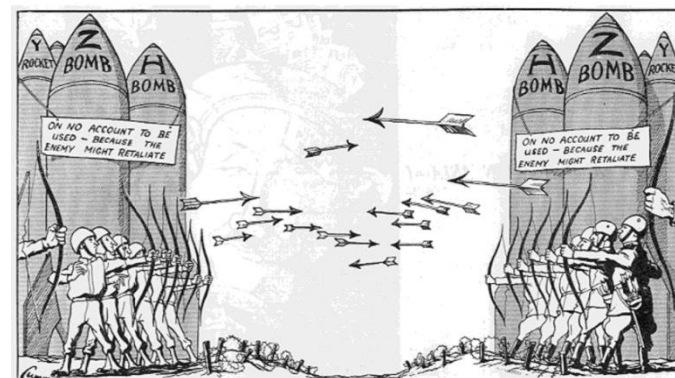
The major turning point in the history of industrial fluoroorganic chemistry was the beginning of the Manhattan Project for development of nuclear weapons in 1941.

The Manhattan Project triggered the need for highly resistant materials, lubricants, coolants and the development of technology for handling extremely corrosive fluoroorganic compounds

After 1945, with the beginning of the Cold War, various defense programs provided a constant driving force for further development of the chemistry and use of organofluorine compounds.

In the 1950s and 60s more civilian applications of fluorinated pharmaceuticals and materials moved into the forefront

The Cold War 1945-1991

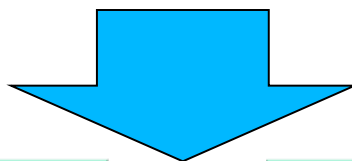


Fluorinated organic compound: Background history

Commercial Uses

Fluorinated surfactants can lower aqueous surface tension to less than 16 dynes/cm and function at very low concentrations (e.g., 100–500 mg/L or parts-per-million, ppm). They are effective in basic/acidic aqueous media and in organic solvents

- superior wetting, spreading, and leveling properties for all types of surfaces.* They give uniform film formation of coatings and eliminates pinholes and craters, even when applied to unclean surfaces
- extremely stable both chemically and thermally.* Some of them are stable even in hot chromic acid, concentrated sulfuric acid or hydrofluoric acid



- Aqueous Film-Forming Foams*
- Enhanced Oil Recovery*
- Coatings*
- Electroplating and Electrowinning*
- Electronics*

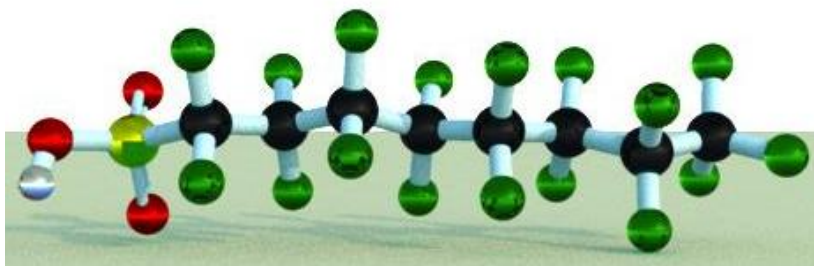
- Paper*
- Mining*
- Photographic Films*
- Fluoropolymer Polymerization Aid*
- Pesticide Application*

Fluorinated organic compound: Background history

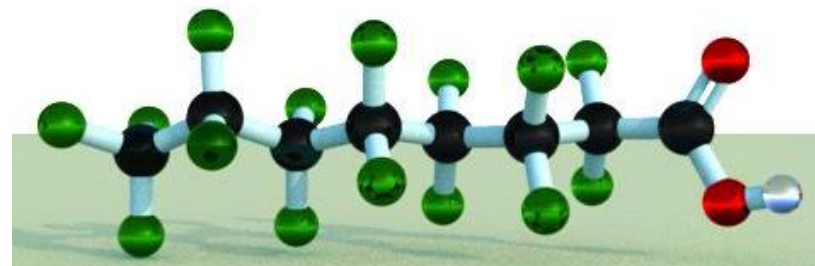
Fluorinated surfactants have been commercially available since the 1950s.

The first available were perfluoroalkyl sulfonates (e.g., perfluorooctane sulfonate, $C_8F_{15}SO_3$, PFOS) and perfluoroalkyl carboxylic acids (e.g., perfluorooctanoic acid, $C_7F_{15}COOH$, PFOA) manufactured using the electrochemical fluorination (ECF) process.

PFOS



PFOA



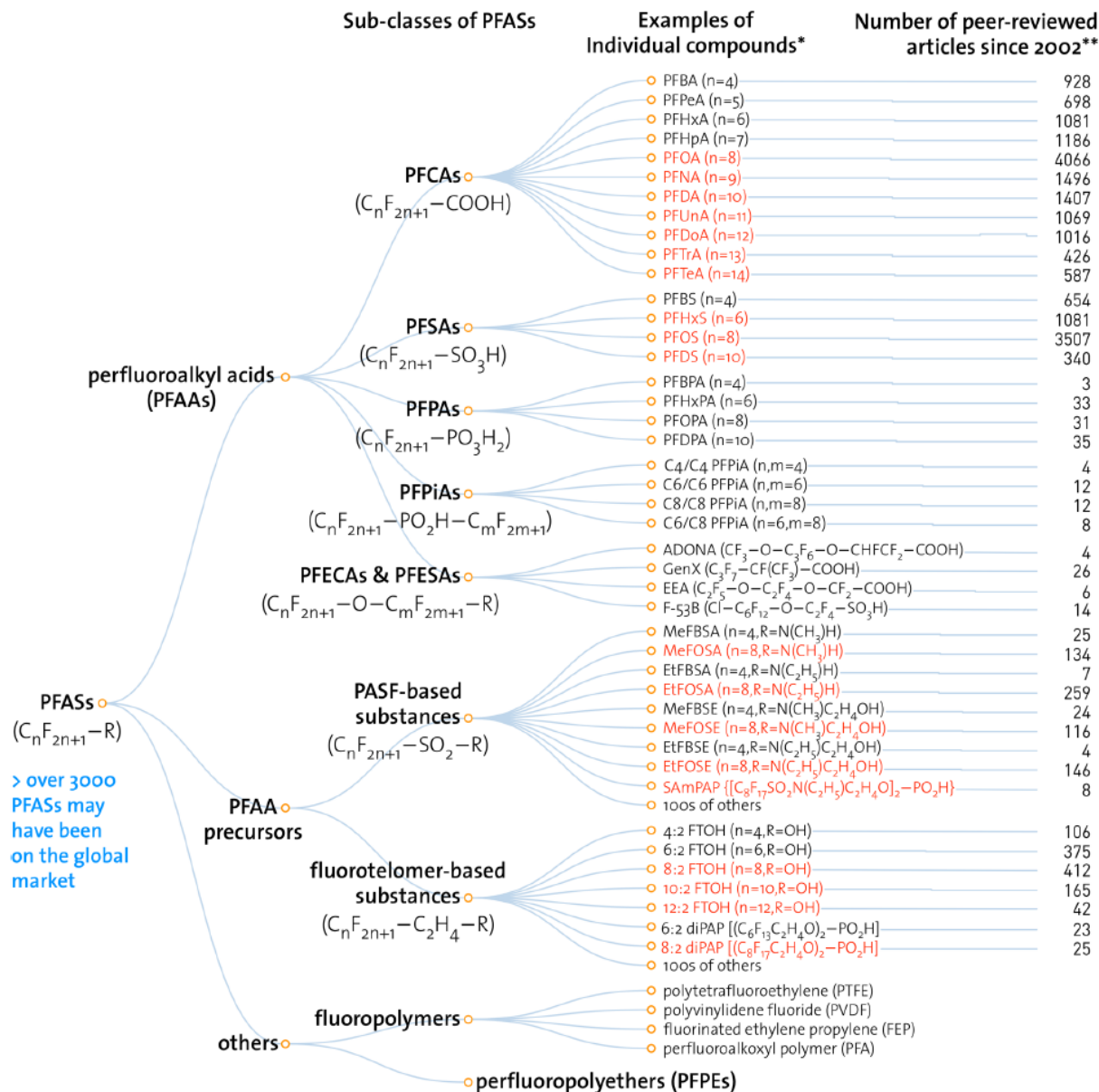
Perfluoroalkyl acids

General name	Acronym	Structure
Perfluoroalkyl sulfonic acid	PFSA	$F(CF_2)_nSO_3H$
Perfluoroalkyl carboxylic acid	PFCA	$F(CF_2)_nCO_2H$
Perfluoroalkyl phosphonic acid	PFPA	$F(CF_2)_nP(=O)(OH)_2$
Perfluoroalkyl phosphinic acid	PFPIA	$F(CF_2)_nP(=O)(OH)$

Fluorinated organic compound: Background history

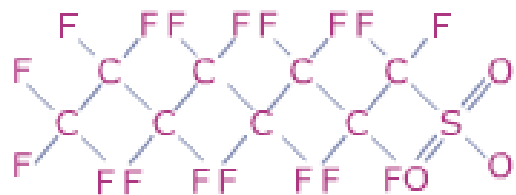
1. PFAs in **RED** are those that have been restricted under national/regional/global regulatory or voluntary frameworks with or without specific exemptions (for details, see OECD (UNEP 2015), Risk reductions approaches for PFASs.<http://oe.cd/1AN>)

2. The numbers of articles (related to all aspects of research) were retrieved from SciFinder on Nov.1, 2016

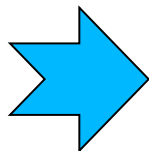


Perfluorinated organic compound: Environmental Regulatory Framework: Elements

IN EUROPEAN UNION



PFOS (Perfluorooctane sulfonate)

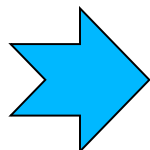


is classified as

- as POP's (Reg. 757/2010) after PFOS was added to the Annex B of the Stockholm Convention in 2009
- Priority Substance for Water (Dir 2013/39/UE)
- Substance with Restriction limit as reported in Reg(EU) No 757/2010 amending Reg (CE) 850/2004



PFOA (Perfluorooctanoic acid)



is classified as

- Candidate in the SVHC List (Substances of Very High Concern) after MSC (Member States Committee) identified in June 2013 PFOA as PBT.
- Substance with Restriction limits as reported in Reg. (UE) 2017/1000 (entry 68)

Perfluorinated organic compound: Environmental Regulatory Framework: Elements

DIRECTIVE 2013/39 of August 2013

COMMISSION DIRECTIVE 2009/90/EC of 31 July 2009

Article 4

Minimum performance criteria for methods of analysis

Member States shall ensure that the minimum performance criteria for all methods of analysis applied are based on an uncertainty of measurement of 50 % or below ($k = 2$) estimated at the level of relevant environmental quality standards and a **limit of quantification equal or below a value of 30 % of the relevant environmental quality standards.**

**Environmental Quality
PAH**

(1)	(2)	(3)					
No	Name of substance	CA number	<p>Requested LOQ = $2,1 \times 10^{-4} \mu\text{g/L}$ Requested LOQ = $3 \mu\text{g/Kg}$</p>				
			$\mu\text{g/L}$	$\mu\text{g/L}$	$\mu\text{g/L}$	$\mu\text{g/L}$	$\mu\text{g/Kg wet weight}$
(35)	Perfluorooctane sulfonic acid and its derivatives (PFOS)	1763-23-1	$6,5 \times 10^{-4}$	$1,3 \times 10^{-4}$	36	7.2	9.1

AA: annual average.

MAC: maximum allowable concentration.

Solid Matrix PFAS Methods

	ASTM D7968	EPA-821-R- 11-007	537Ms / DoD
MATRIX	Soils	Sludge, Biosolids	Soils, Sediments, Biosolids, Tissues, etc
RL (ng/g)	0.025 - 0.75	0.25 - 10	<i>var</i>
PREPARATION	SLE (rotator) centrifuge, filter	digestion, incubation, SLE (shake), SPE	var
CLEAN-UP	Filtration	SPE WAX + filtration	var

SLE = Solid-Liquid Extraction

SPE = Solid Phase Extraction

WAX = Weak Anion Exchange

Recent developments in Solid Matrix PFAS Methods

QuEChERS is considered accurate and highly productive at ultra trace levels¹⁰. Yet, for the analysis of PFAS in food, this method is not widely applied compared to the straightforward SLE and IPE methods.

Recently, a one step QuEChERS extraction and purification was found to be successful.



FDA U.S. FOOD & DRUG
ADMINISTRATION

FDA Foods Program Compendium of Analytical Laboratory Methods: Chemical Analytical Manual (CAM)

METHOD NUMBER: C-010.01

POSTING DATE: 11/01/2019

POSTING EXPIRATION DATE: 10/31/2021

PROGRAM AREA: Chemical Contaminants

Determination of 16 Perfluoroalkyl and Polyfluoroalkyl
Substances (PFAS) in Food using Liquid Chromatography-
Tandem Mass Spectrometry (LC-MS/MS)

Version 2019 (2019)

Author: Susan Genualdi and Lowri deJager

CFSAN/ORS reviewers: Tim Begley, Gregory Noonan

GLOSSARY

Analytical Methods

PAPER



Cite this: *Anal. Methods*, 2018, 10, 5715

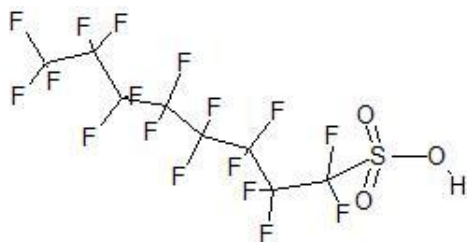


[View Article Online](#)
[View Journal](#) | [View Issue](#)

Simultaneous determination of legacy and emerging per- and polyfluoroalkyl substances in fish by QuEChERS coupled with ultrahigh performance liquid chromatography tandem mass spectrometry†

Yan Gao,  Qinghe Zhang,* Xiaomin Li,  Xiuqin Li and Hongmei Li

Perfluorinated organic compound: Challenges for the Analysis: Choice of Mass Detector

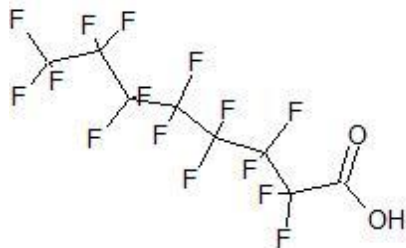


PFOS

 ESI neg $[C_8F_{17}SO_3]^-$

 mass=498.93022

 nominal=498



PFOA

 ESI neg $[C_8F_{15}O_2]^-$

 mass=412.96643

 nominal=413

The main measurement technique for Perfluorinated compounds is liquid chromatography coupled to mass spectrometry after negative electrospray ionisation.

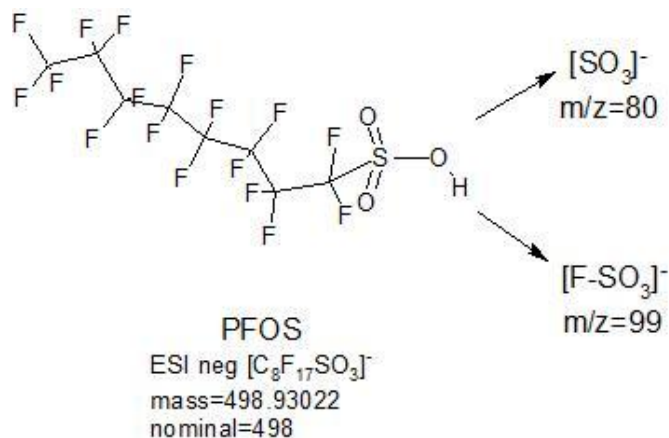


In ESI (negative mode)

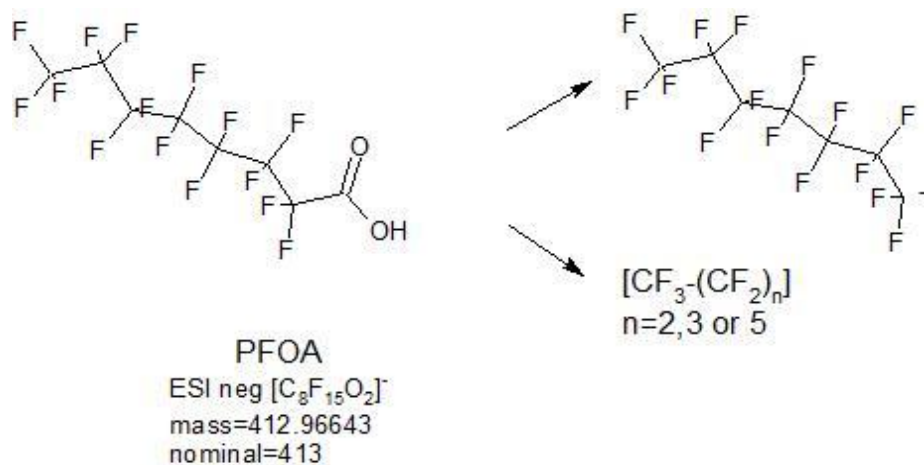
The pseudomolecular ions $[M-H]^-$ is observed as the main generated ionic species.

Perfluorocarboxylic acid, such as PFOA, exhibit in source fragmentation with loss of CO_2

Perfluorinated organic compound: Challenges for the Analysis: Choice of Mass Detector: behaviour in MSMS mode



Loss of the hydrophilic sulfonate group was observed, leading to $[SO_3]$ (m/z 80) and/or $[FSO_3]$ (m/z 99) ions. However, these fragment ions remained of poor intensity and limited specificity. Example of interferent in biological samples: taurodeoxycholic acid (TDCA)



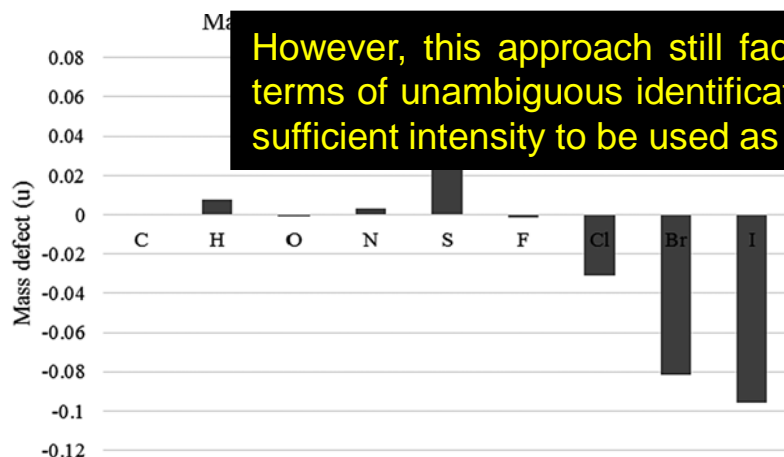
For PFOA (Figure 2b), the observed fragmentation appeared slightly more effective, with the loss of CO_2 and subsequent fragmentation on the alkyl chain leading to $[CF_3-(CF_2)_n]$ ions where n equals 2, 3 or 5.

Perfluorinated organic compound: Challenges for the Analysis: **Choice of Mass Detector : behaviour in HRMS mode**

The relative atomic mass of fluorine is 18.9984 and this value is less than the nominal mass (19).

In the case of hydrogen, the mass is larger than the nominal mass (1.008 vs 1)

Highly fluorinated compounds will therefore have lower monoisotopic masses than their respective nominal mass, in respect to compounds with only C–H bonds



However, this approach still faces some difficulties in fulfilling strict regulatory criteria in terms of unambiguous identification of the target analytes as only one ion is available with sufficient intensity to be used as a diagnostic signal.

The use of mass defect plots for the identification of (novel) halogenated contaminants in the environment

Karl J. Jobst • Li Shen • Eric J. Reiner •
Vince Y. Taguchi • Paul A. Helm • Robert McCrindle •
Sean Backus

These properties can be very useful for the identification of PFASs with high resolution instruments capable of measuring monoisotopic mass.

Instrumental Choice



Orbitrap Exactive HCD

PFOS: **Biota: Pool of species**

According to ISPRA guidelines, the species selected for the monitoring of PFAS, were:



Chelon labrosus



Liza aurata



Liza ramada



Liza saliens



Mugil cephalus

Sea and transiction waters:

All trophic levels betwee 2,3 and 3,5



Barbus plebejus



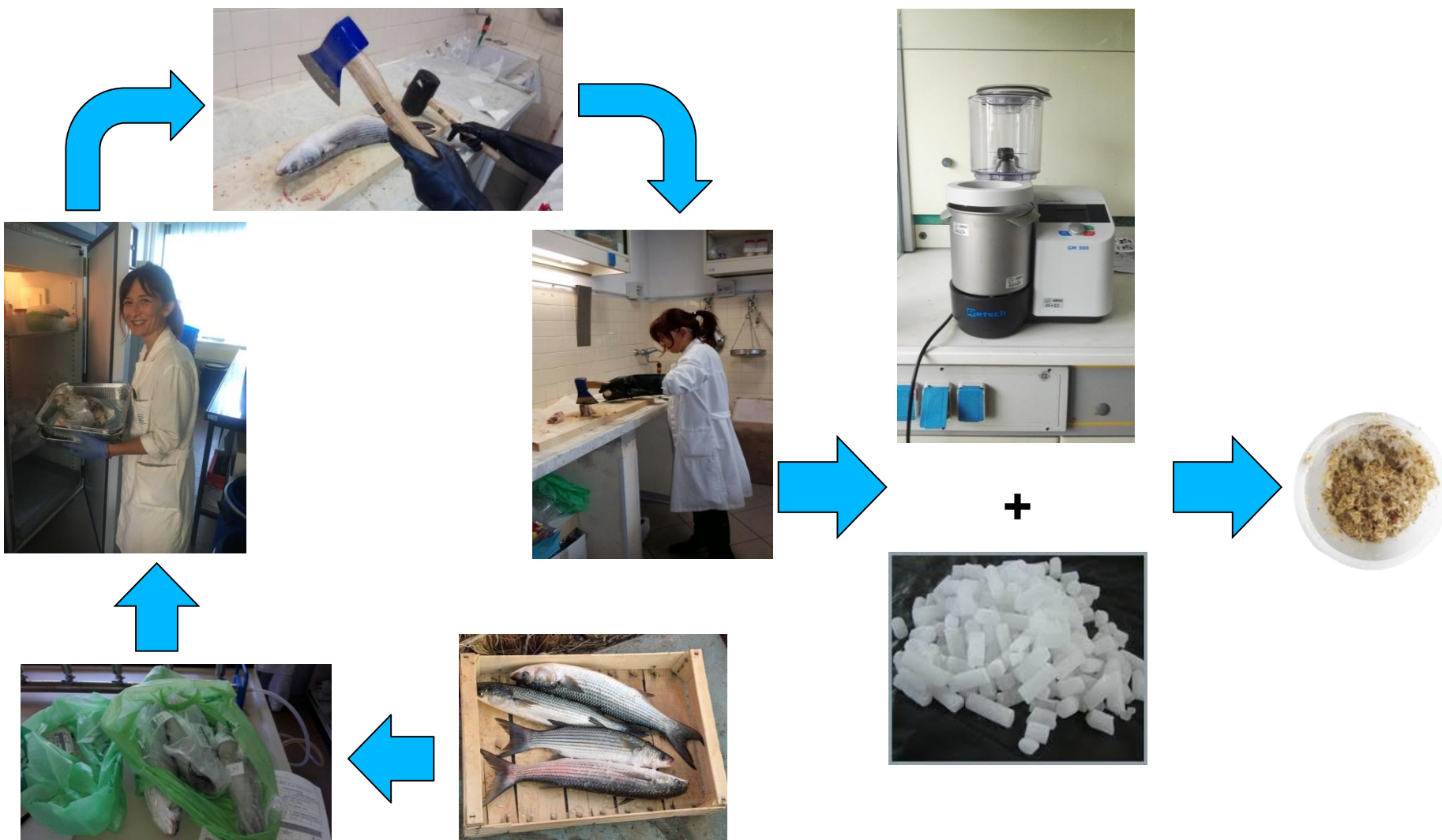
Leuciscus cephalus



Salmo trutta

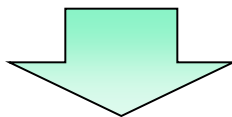
River and lakes

Fluorinated organic compound: Sample Treatment: Fish



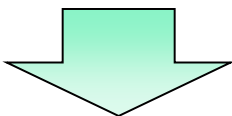
Analysis of PFOS in Fish: QUECHERs Extraction

Addition of 30 μ l of 200 ng/ml methanolic solution of Extraction ILS ($^{13}\text{C}_8$ -PFOS) to 2 g of fish omogeneate.



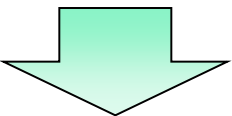
equilibrate for 24 hours at -18°C

Addition of 2 ml of water and 1 ceramic homogeneizer,



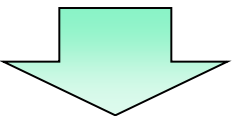
vortex agitation for 1 minute

Addition of 10 ml of acetonitrile



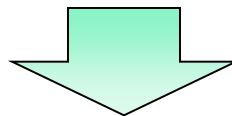
vortex agitation for 1 minute

Addition of 1 pouche of reagent mix for QUECHERs extraction compliance to EN15662



vortex agitation for 1 minute

Centrifugation and collection of liquid phase



Withdrawal of 1,2 ml of solution and addition of 0,3ml of water

412 ANASTASSIADES ET AL., JOURNAL OF AOAC INTERNATIONAL VOL. 86, NO. 2, 2003

RESIDUES AND TRACE ELEMENTS

Fast and Easy Multiresidue Method Employing Acetonitrile Extraction/Partitioning and "Dispersive Solid-Phase Extraction" for the Determination of Pesticide Residues in Produce

MICHELANGELO ANASTASSIADES¹ and STEVEN J. LEHOTAY²

¹U.S. Department of Agriculture, Agricultural Research Service, Eastern Regional Research Center, 600 E. Mermaid Ln, Wyndmoor, PA 19038

²DARINKA STAINBAHR

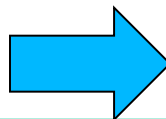
Public Health Institute, Environmental Protection Institute, Prvomajska 1, 2000 Maribor, Slovenia

FRANK J. SCHENCK

U.S. Food and Drug Administration, Office of Regulatory Affairs, Southeastern Regional Laboratory, 60 Eighth St, Atlanta, GA 30309

pH Adjustment in QUECHERS Extraction Step

Various Buffers tested



Compromise: Citrate Buffer

4g Magnesium sulphate anhydrous,

- 1 g Sodium chloride (not essential but kept for **better selectivity**),
- **1 g Trisodium citrate dihydrate and**
- **0.5 g Disodium hydrogencitrate sesquihydrate**

Merits:

- ✓ **Good recoveries even for very acidic pesticides (dicamba)**
- ✓ **Good recoveries for base- and acid-sensitive pesticides**
- ✓ **Improved Selectivity (less co-extractives from **acidic samples**)**
- ✓ **No negative effect on PSA cleanup (unlike Acetate Buffer)**

Impact of Lipids on Workflow



• **Inability to meet detection criteria**

- Longer method development time
- Method troubleshooting
- Variability depending on matrix
- QC/LOQ issues/reruns
- **Indeterminate time**

• **Data variability**

- High RSDs
- Data accuracy
- Variability depending on matrix
- **Reruns, data analysis time**

• **Mass Spec Cleanliness**

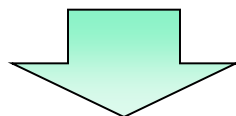
- Cleaning the source
- Cleaning/replacing capillary
- Liner Lifetime/Inlet issues
- Establishing vacuum levels
- Retuning
- Any additional troubleshooting
- **4 hours - > 1day**

• **Lipid Build Up on Column**

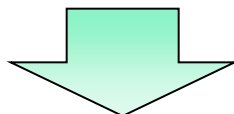
- Column longevity / GC column cutting
- Back pressure
- Column flushing / equilibration
- **Approximately 2 hours**

Analysis of PFOS in Fish: Clean-up

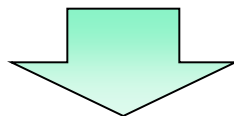
Load 1,5 ml of dilute extraction solution on the top of an Agilent EMR Captiva 3 mL, 300 mg



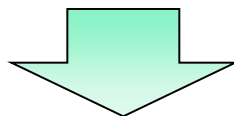
Allow extract flow under gravity and collect into a polypropylene tube



Load 300 μ L of Acetonitrile/Water = 80/20, flow under gravity and collect with the previous fraction



Drain with positive pressure (Syringe) and vortex the combined eluates



Take 300 μ L of combined eluates, put into a polypropylene vial and add 4,5 μ L of ILS injection solution (1,2,3,4- $^{13}\text{C}_4\text{C}_4_4$ -PFOS , 200 ng/ml: 0,9 ng, approximately 3 ng/ml)

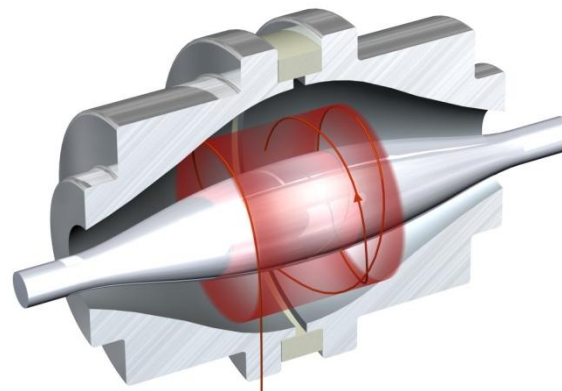
PFOS in Fish: Analytical condition

UHPLC: Thermo Accela 1250 pump

- *TRAP column: 50x2.1 mm (1.9 μ m) Waters Fusion (between pump and injection valve)*
- Analytical column : Phenomenex Kinetex F5, 100 mm, 3 mm, 2,6 μ m
- Mobile Phase: [A] 5mM NH_4OOCH +2%MeOH + 0.025% HCOOH ; [B] MeOH 85%+iPrOH 15%
- Gradient from 90/10 to 10/90 in 20 min., Flow: 400 $\mu\text{L}/\text{min}$
- Column Temperature: 40 $^{\circ}\text{C}$

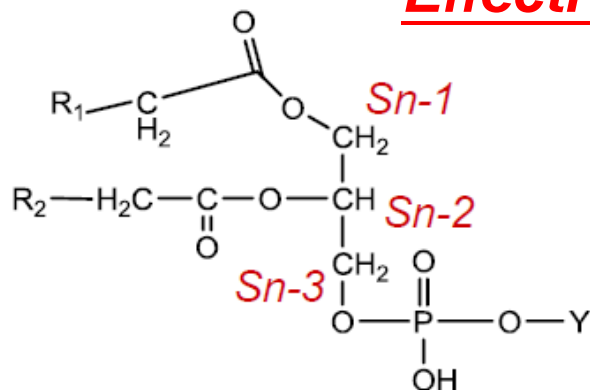
Thermo Orbitrap Exactive

- ESI negative
- Full scan, Resolution 50000, accuracy 5 ppm

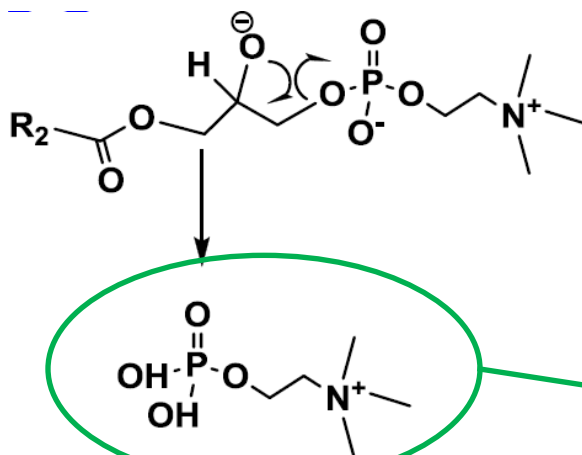


Analyte	Deprotonated ion	Target ion
Perfluorooctanesulfonic acid	$\text{C}_8\text{F}_{17}\text{O}_3\text{S}$	498.93022
$^{13}\text{C}_8$ Perfluorooctanesulfonic acid (ILS)	$^{13}\text{C}_8\text{F}_{17}\text{O}_3\text{S}$	506.95706
1,2,3,4 $^{13}\text{C}_8$ Perfluorooctanesulfonic acid (ILS)	1,2,3,4 $^{-13}\text{C}_4\text{C}_4\text{F}_{17}\text{O}_3\text{S}$	502.94364

Effectiveness of Clean Up Step



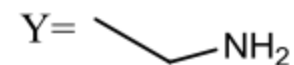
Phospholipid: general structure



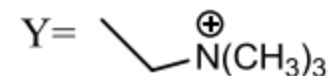
$m/z=184,07332$

Phosphocoline- H^+

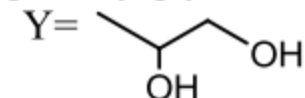
Phosphatidylethanolamine (PE)



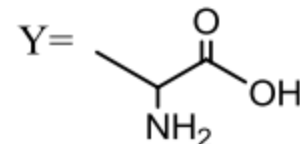
Phosphatidylcholine (PC)



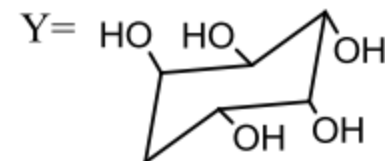
Phosphatidylglycerol (PG)



Phosphatidylserine (PS)



Phosphatidylinositol (PI)



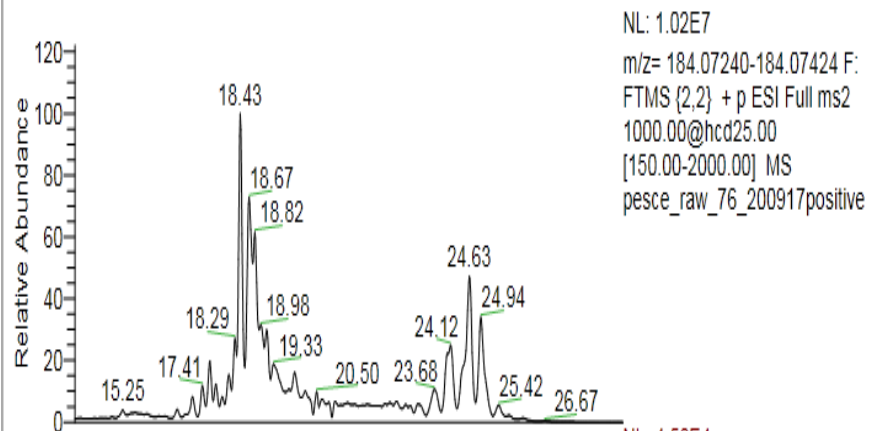
Phosphatidylcholine: Typical fragmentation

Effectiveness of Clean Up Step

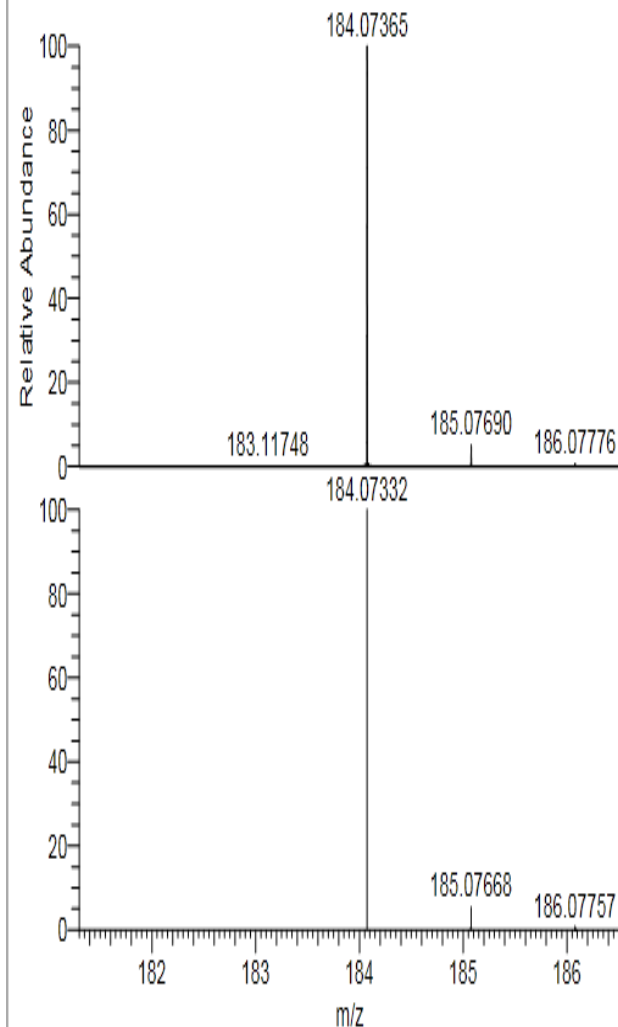
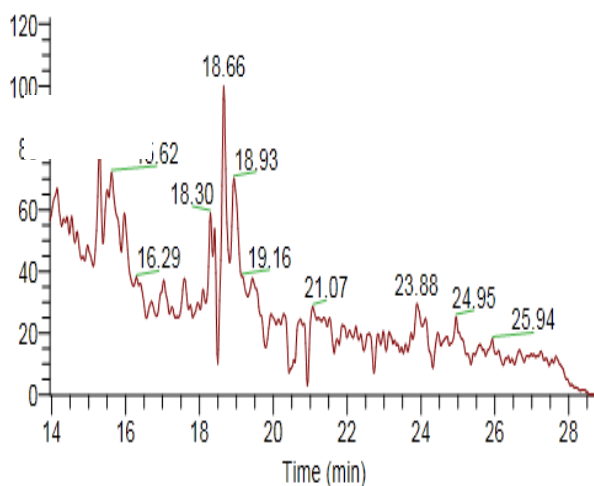
pesce_raw_76_200917positive

09/17/20 18:02:00

RT: 13.95 - 28.79 SM: 9G



NL: 4.56E4
m/z= 184.07240-184.07424 F:
FTMS {2,2} + p ESI Full ms2
1000.00@hcd25.00
[150.00-2000.00] MS
pesce_captiva_76_200917_positiv
e_1



NL:
7.90E6
pesce_raw_76_200917positive
#2193 RT: 18.40 AV: 1 F:
FTMS {2,2} + p ESI Full ms2
1000.00@hcd25.00
[150.00-2000.00]

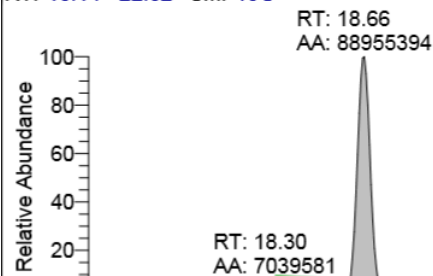
NL:
9.33E5
C₅H₁₅PO₄N:
C₅H₁₅P₁O₄N₁
pa Chrg 1

Effectiveness of Clean Up Step

Pesce_RAW_77_200917_positive

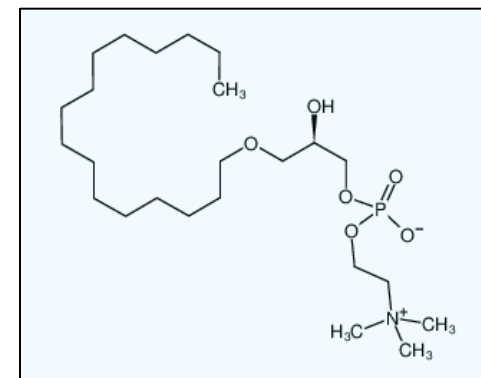
09/17/20 18:41:36

RT: 16.14 - 22.62 SM: 15G



NL: 1.06E7
m/z= 496.33729-496.34225
F: FTMS {2,1} + p ESI Full
ms [100.00-2000.00] MS
ICIS
Pesce_RAW_77_200917_po
sitive

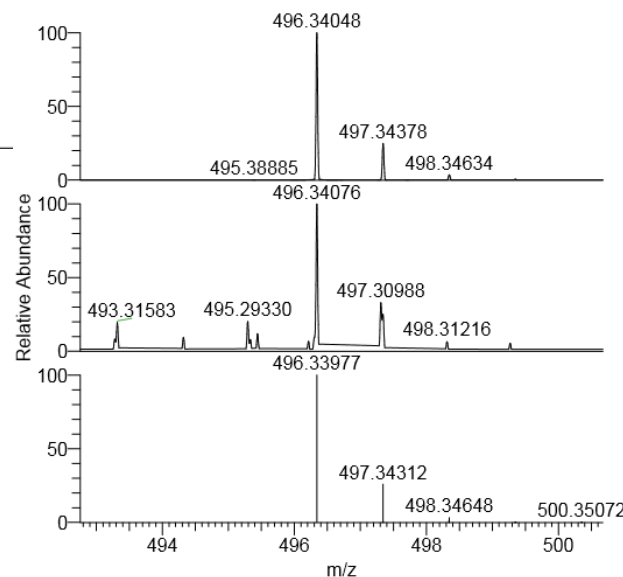
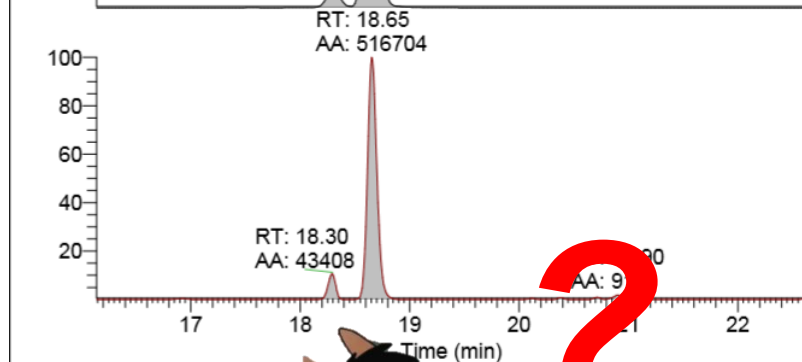
NL: 8.61E4
m/z= 496.33729-496.34225
MS ICIS
pesce_captiva_77_200917_p
ositive_1



Lysolecithin

Lysophosphatidylcholine (LPC)

2-Hydroxy-3-(palmitoyloxy)propyl 2-(trimethylammonio)ethyl phosphate



NL:
4.05E6
Pesce_RAW_77_200917_positive#22
29 RT: 18.69 AV: 1 T: FTMS {2,1} +
p ESI Full lock ms [100.00-2000.00]

NL:
1.07E4
pesce_captiva_77_200917_positive_1
#2242 RT: 18.65 AV: 1 T: FTMS
{2,2} + p ESI Full ms2
1000.00@hcd25.00 [150.00-2000.00]

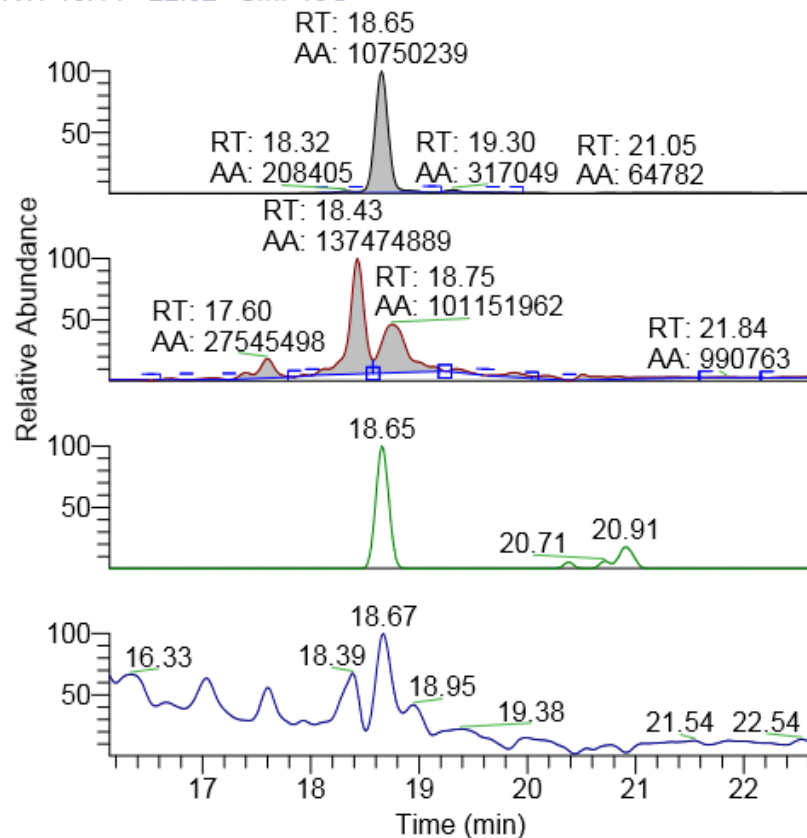
NL:
7.52E5
C₂₄H₅₀NO₇P + H:
C₂₄H₅₁N₁O₇P₁
pa Chrg 1

Effectiveness of Clean Up Step

pesce_captiva_77_200917_positive_1

09/17/20 16:03:10

RT: 16.14 - 22.62 SM: 15G

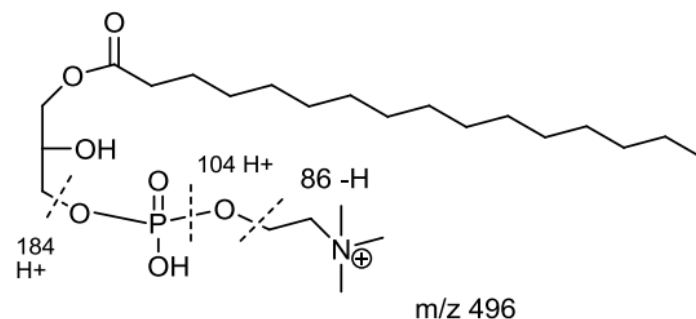


NL: 1.26E6
m/z= 496.33729-496.34225 F: FTMS {2,2} + p ESI
Full ms2 1000.00@hcd25.00 [150.00-2000.00] MS
ICIS Pesce_RAW_77_200917_positive

NL: 1.51E7
m/z= 184.07228-184.07412 F: FTMS {2,2} + p ESI
Full ms2 1000.00@hcd25.00 [150.00-2000.00] MS
ICIS Pesce_RAW_77_200917_positive

NL: 1.29E4
m/z= 496.33729-496.34225 F: FTMS {2,2} + p ESI
Full ms2 1000.00@hcd25.00 [150.00-2000.00] MS
pesce_captiva_77_200917_positive_1

NL: 3.36E4
m/z= 184.07228-184.07412 F: FTMS {2,2} + p ESI
Full ms2 1000.00@hcd25.00 [150.00-2000.00] MS
pesce_captiva_77_200917_positive_1

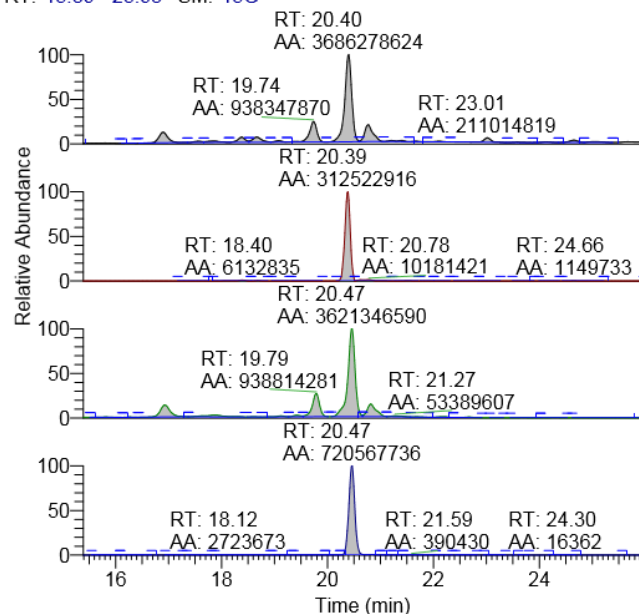


Effectiveness of Clean Up Step

pesce_raw_77_200917_negative

09/18/20 00:00:21

RT: 15.39 - 25.93 SM: 15G

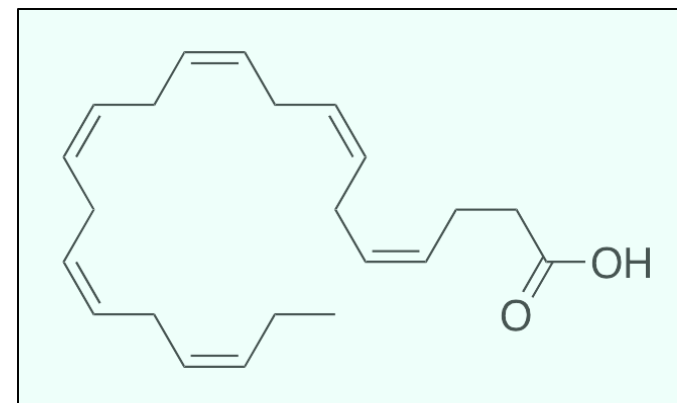


NL: 4.03E8
TIC F: FTMS {2,1} - p ESI Full ms
[100.00-2000.00] MS ICIS
pesce_raw_77_200917_negative

NL: 4.34E7
m/z= 327.23022-327.23350 F: FTMS {2,1} - p
ESI Full ms [100.00-2000.00] MS ICIS
pesce_raw_77_200917_negative

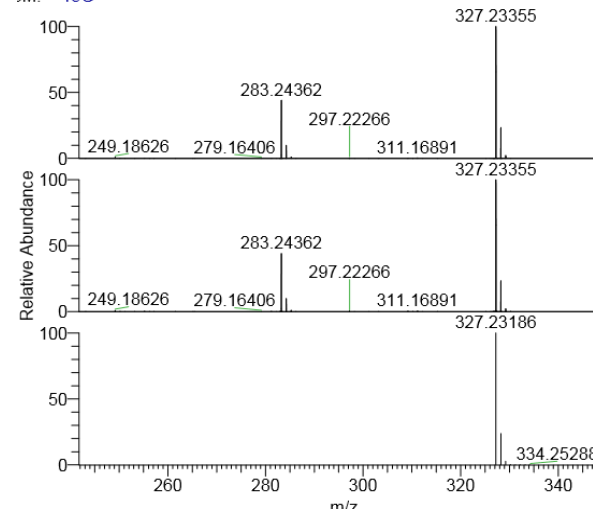
NL: 3.84E8
TIC F: FTMS {2,1} - p ESI Full ms
[100.00-2000.00] MS ICIS
Pesce_captiva_77_200917_negative

NL: 9.04E7
m/z= 327.23022-327.23350 F: FTMS {2,1} - p
ESI Full ms [100.00-2000.00] MS ICIS
Pesce_captiva_77_200917_negative



Docosahexaenoic acid (DHA)

SM: 15G

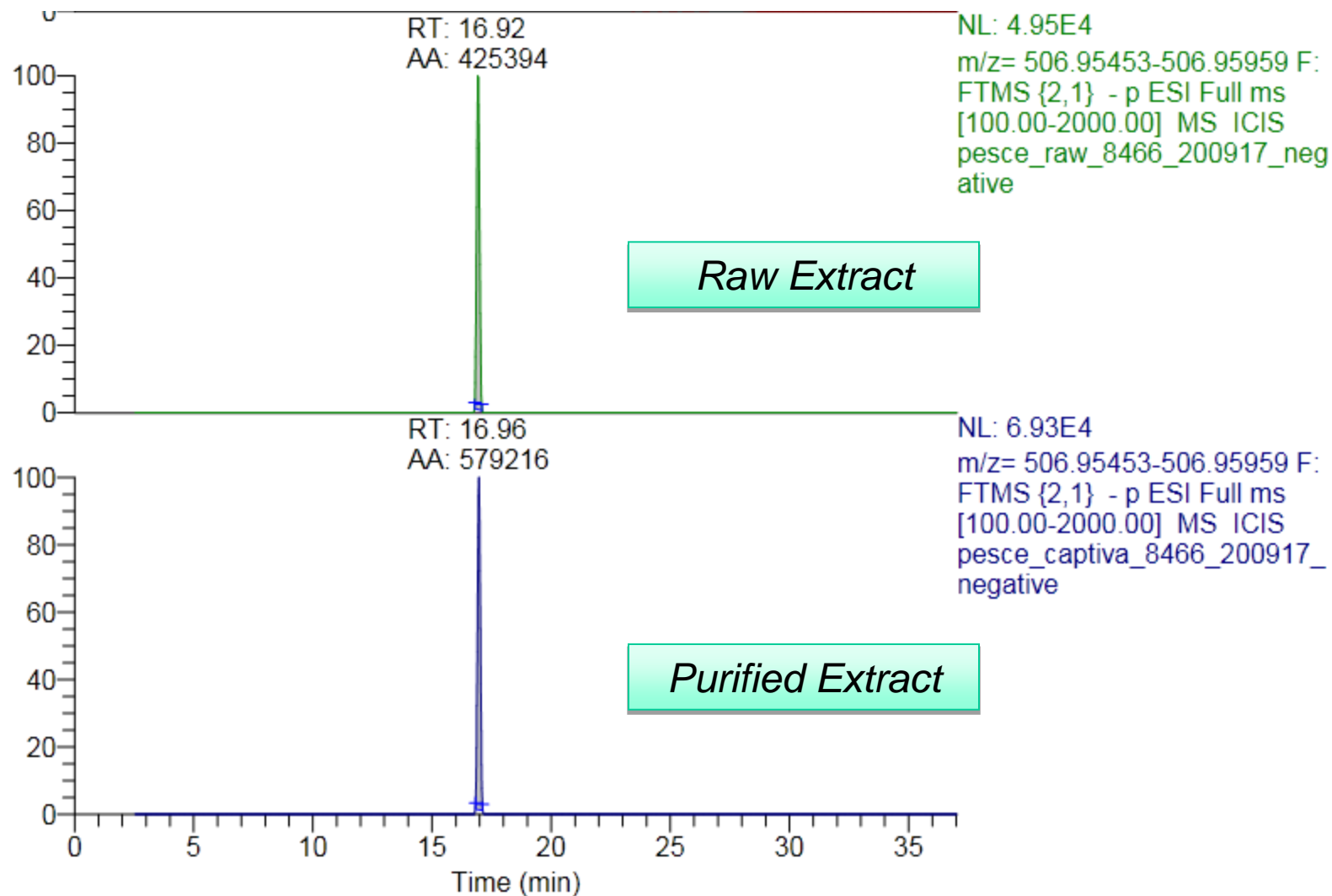


NL: 1.14E8
Pesce_captiva_77_200917_neg
ative#2280 RT: 20.45 AV: 1 T:
FTMS {2,1} - p ESI Full ms
[100.00-2000.00]

NL: 1.14E8
Pesce_captiva_77_200917_neg
ative#2280 RT: 20.45 AV: 1 T:
FTMS {2,1} - p ESI Full ms
[100.00-2000.00]

NL: 7.83E5
C₂₂H₃₁O₂
C₂₂H₃₁O₂
pa Chrg 1

Effectiveness of Clean Up Step



Validation Study



Pikeperch, is a species of ray-finned fish from the family Percidae. It is found in freshwater and brackish habitats in western Eurasia.

	MASS FRACTION	
	Certified value (ng/g)	Uncertainty (ng/g)
Linear perfluorooctane sulfonate (L-PFOS) ⁽¹⁾	16 ⁽²⁾	1,7 ⁽²⁾

1) As defined by using liquid chromatography mass spectrometry.

2) Unweighted mean value of the means of accepted sets of data, each set being obtained in a different laboratory with a method of determination including liquid chromatography mass spectrometry. Sulfonates are expressed on an anion basis. The certified/ values and their uncertainties are traceable to the International System of Units (SI).

3) The uncertainty of the certified / indicative value is the expanded uncertainty with a coverage factor $k = 2$ corresponding to a level of confidence of about 95 % estimated in accordance with ISO/IEC Guide 98-3, Guide to the Expression of Uncertainty in Measurement (GUM:1995), ISO, 2008

Validation Study

The IRMM 427 (certified concentration around 16 ng/g) was analyzed compliance to ARPAT-MICAVL015 during the period between 7/01/2020 - 24/02/2020 for a total of 19 times with different analysts.

Application Note 1



Comparison of a measurement result with the certified value

January 2010

The comparison of a measurement result on a certified reference material with the certified value is explained. The method compares the difference between the certified and measured values with its uncertainty, i.e. the combined uncertainty of certified and measured value. Guidance on how to determine the standard uncertainties of certified values as well as standard uncertainties of measurement results is given.

Author: Thomas Linsinger
European Commission - Joint Research Centre
Institute for Reference Materials and
Measurements (IRMM)
Retieseweg 111, 2440 Geel, Belgium
Email: thomas.linsinger@ec.europa.eu
www.erm-crm.org

A second level of concentration (about 2 ng/g) was investigated through the participation to the Inter-agency exercise during summer of 2019

Validation Study

After the measurement of a CRM the absolute difference between the mean measurement and the certified value is

To evaluate method performance,

Δ_m is compared with U_Δ :

If

$$\Delta_m \leq U_\Delta$$

Then

there is no significant difference between the measurement result and the certified value.

$$U_{CRM} = U_{Certified Value} / 2$$

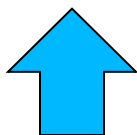
3) The uncertainty of the certified value is the expanded uncertainty with a coverage factor $k = 2$ corresponding to a level of confidence of approximately 95 % estimated in accordance with ISO/IEC Guide 98-3, Guide to the Expression of Uncertainty in Measurement (GUM:1995), ISO, 2008.

Validation Study

Choice of quantification Mode

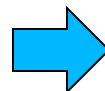
$$C_x = (A_x / A_{xILS}) \times C_{xILS}$$

Barbara Maier and Michael Vogeser, Clin Chem Lab Med 2012; 51(4): 833–837



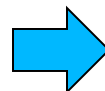
*Significant difference between
the measurement result and
the certified value*

SIDA (Stable Isotope Dilution Analysis)



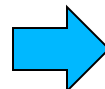
NO

Pure Solvent Standard



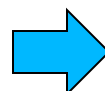
YES

Matrix Standard



YES

Processed Solvent Standard



NO

Validation Study

Partecipation of Laboratory in n Proficiency Test

Uncertainty

$$\bar{u}_{Cref} = \frac{\sum u_{Cref,j}}{n}$$



Average of Uncertainties of n PT

$$d_i = x_i - C_{ref}$$



Difference between the measurement results and
assigned value of the *i*th sample of the
interlaboratory comparison
(Laboratory Bias in *i*th PT)

$$D_{iRel} = d_i / C_{ref}$$



Relative Difference of the *i*th sample of the
interlaboratory comparison

$$D_{ms,rel} = \sqrt{\frac{\sum D_{i,rel}^2}{N}}$$



Root mean square of the relative
difference
(Standard Deviation of Relative Bias)

Validation Study

	Pure Solvent Standard	Processed Standard	Matrix Standard	SIDA
Extended Uncertainty	41.1	44.4	42.2	38.4

Sist
e

Recent trends in inter-laboratory precision at ppb and sub-ppb concentrations in relation to fitness for purpose criteria in proficiency testing

Michael Thompson

School of Biological and Chemical Sciences, Birkbeck College (University of London),
Gordon House, 29 Gordon Square, London, UK WC1H 0PP

Received 12th January 2000, Accepted 31st January 2000

The
Analyst
COMMUNICATION

⇒ Random
error

$$U_{c,rel} = k$$

$$\sigma = \begin{cases} 0.22c & \text{if } c < 1.2 \times 10^{-7} \\ 0.02c^{0.8495} & \text{if } 1.2 \times 10^{-7} \leq c \leq 0.138 \\ 0.01c^{0.5} & \text{if } c > 0.138 \end{cases}$$

ainty

m Manual 206/1, Rev 2015

Validation Study

Inter-Agency Exercise

Laboratory	A	B	C	D	E	G
	2,37	1,90	2,53	1,00	2,52	3,95
	2,18	1,80	2,42	1,30	2,98	3,46
	2,38	2,20	2,56	1,10	3,65	3,66
Mean	2,31	1,97	2,50	1,13	3,05	3,69
Dev. St.	0,11	0,21	0,07	0,15	0,57	0,25
CV%	4,88	10,58	2,94	13,48	18,63	6,68

1° G = 3 LAB	1,00	Mean	1,55	2,37
	1,10	Dev. St.	0,48	2,18
	1,30	CV%	31,00	2,38
	1,80			2,52
	1,90			2,98
2G = 4 Lab	2,18			3,65
	2,20	Mean	2,40	1,90
	2,37	Dev. St.	0,12	1,80
	2,38	CV%	5,03	2,20
	2,42			2,53
3° G = 3 Lab	2,52			2,42
	2,53			2,56
	2,56	Mean	3,38	
	2,98	Dev. St.	0,51	
	3,46	CV%	15,19	
	3,65			
	3,66			
	3,95			

Validation Study

Summary

Quantification Mode: SIDA

Range:

2 - 16 $\mu\text{g/Kg}$

Uncertainty (conc. equal or higher than 9.2 $\mu\text{g/kg}$):

50%

Uncertainty (conc. less than 9,2 $\mu\text{g/kg}$):

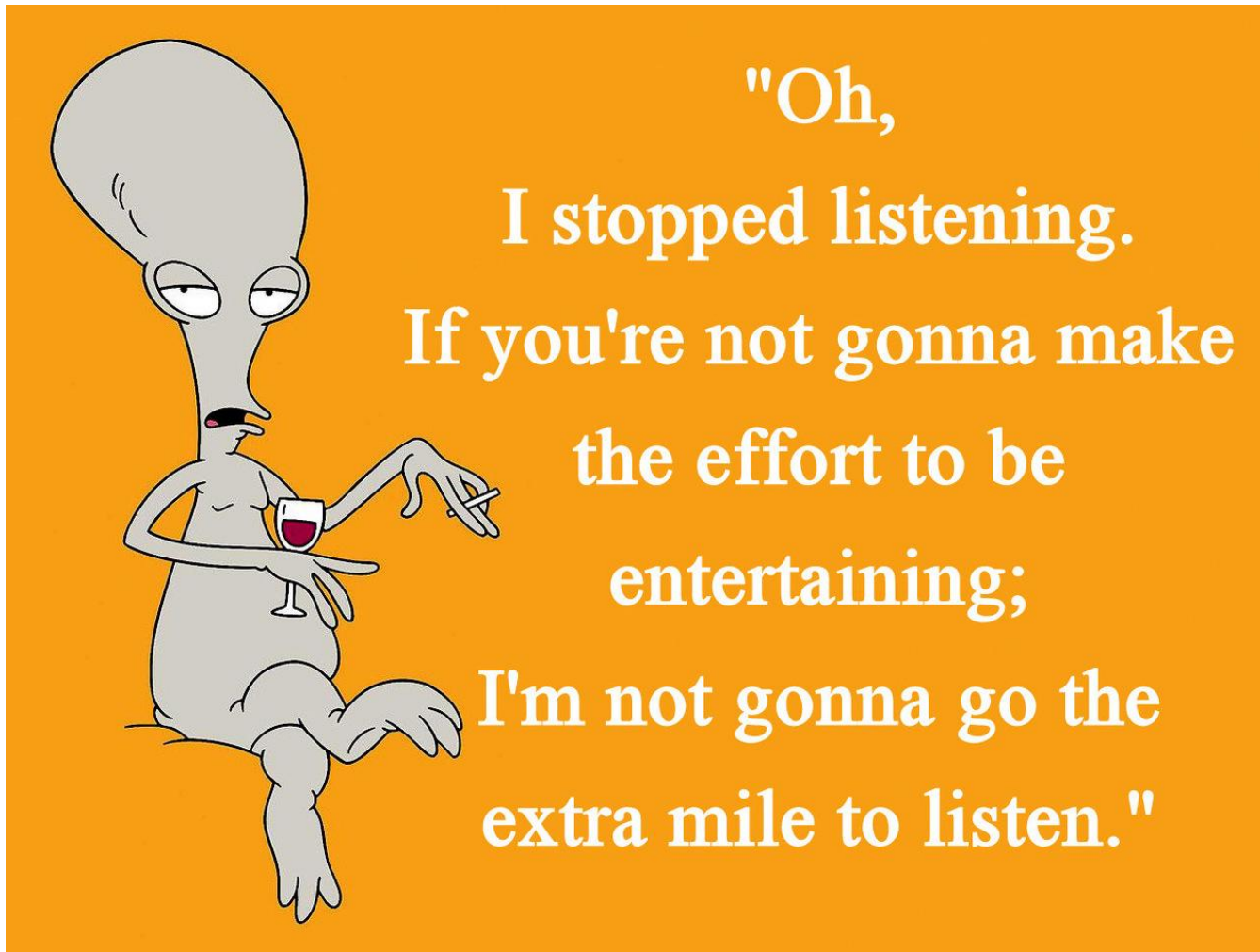
57 %

LOQ:

2 ng/g

Limit of repeatibiliy:

20%



Thanks